



ORIGINAL ARTICLE

Determination of Organophosphorus Pesticide Residues in Cabbages from Bindura Market Place by Solid Phase Extraction and Gas Liquid Chromatography

Shasha D, Gwezere W, Dzomba P, Chayamiti T

¹Chemistry Department, Faculty of science, Bindura University of Science Education, P. Bag 1020, Bindura, Zimbabwe

*Corresponding author: E-mail: pdzomba@gmail.com, pdzomba@buse.ac.zw.

ABSTRACT

The study investigated presence of organophosphate pesticides residues in cabbages sold at Bindura market place. Gas Chromatography-Nitrogen Phosphorus Detector (GC-NPD) coupled to solid phase extraction on florasil column (500mg/8ml) cartridges was used to determine organophosphate pesticides levels. Percentage recovery was above 76%. Limit of detection and quantification was 0.1ng/ml and 2.35ng/ml respectively. Dimethoate was the mostly detected pesticide. The highest level, 0.81 ± 0.02 was obtained from Aerodrome and the lowest level, 0.02 ± 0.00 was recorded for Magobo cabbages. Overall a total of 51 pesticides residues were detected in 180 samples corresponding to detection rate of 28%. The implicated organophosphate pesticides include; acephate, dimethoate; malathion, chlopyrifos-methyl and monocrotophos. All the levels obtained in the present study were far below the maximum recommended safety limits showing that farmers were sticking to recommended waiting times before harvesting their cabbages. Although levels recorded are low however presence of more than one pesticide residue type is worrisome because of a possibility of synergic tendencies. Usually safety limits are for a single pesticide and not for multiple pesticides.

Key words: organophosphorus pesticides, cabbages, solid phase extraction, gas chromatography

Received 12/12/2013 Accepted 19/01/2014

©2014 AELS, INDIA

INTRODUCTION

Organophosphorus pesticides (OPPs) are widely applied in pre and post harvest treatment to control diseases in vegetables such as cabbages [1]. Applied pesticides have been found to persist over the season such that they may appear in vegetables meant for human consumption [2]. Presence of pesticides in vegetables has health implications. They can accumulate in the blood and adipose tissue and reach toxic levels [3,4]. Humans are exposed to organophosphate pesticides via inhalation, absorption through the skin and mostly by ingestion of contaminated vegetables and fruits [5]. Inside the body OPPs bind with enzyme cholinesterase at a synapse and paralyze enzyme activity resulting in elevated acetylcholine [6]. Consequently red blood cells and serum cholinesterase cease to function. Exposure to high levels of OPPs results in acute intoxication characterized by poisoning symptoms such as gastro-intestinal upsets, bronchospasms and urination [7]. In pregnant mothers high level exposure cause spontaneous abortion and fetal death. Previous studies have found Hodgkin's lymphoma in farm employees and cancers in their children as a result of OPPs exposure [8] European Protection Agency (EPA) reports that all OPPs are toxic to the nervous system [9, 10].

Park et al., [11] investigated organophosphate pesticides in agricultural produce and found that they were not free from pesticides. Wen et al., [12] also observed persistence of organophosphate pesticides in agricultural produce. In most developing countries like Zimbabwe there are no strict control measures to regulate the use of pesticides. Although importation and licensing of pesticides is usually regulated by the government selling and buying is often not controlled such that consumers are free to buy and use pesticides at will. Pesticides residues screening on foods is often not mandatory. Due to shrinking economy most people in developing countries ventured into market gardening and most sell their produce at markets such as Bindura Market place. Most of these farm produce would not have passed

through pesticides residue screening regimes. In such a situation researches dedicated to determine pesticide residues levels in such products become most crucial thereby providing the rationale behind this study. Pesticides of interest in this study are the most frequently used; acephate, monocrotophos, dimethoate, malathion and Chlopyriphos-methyl determined by interviewing twenty farmers per each sampling day for five days Table 1. Participation in the survey was voluntary.

Table 1. Mostly applied OPPs in Bindura farming area

Vegetables origin	Pesticide used	Frequency of quote (%)
Musana	acephate	77
	dimethoate	90
Manhenga	Monocrotophos	55
	dimethoate	83
Supa	Dimethoate	33
Madziwa	Dimethoate	67
	malathion	22
Glendale	Malathion	56
	Dimethoate	78
Matepatepa	Dimethoate	34
	Chlopyriphos- methyl	57
Shamva	Chlopyriphos- methyl	88
	Dimethoate	72
Chipadze	Dimethoate	66
	Chlopyriphos- methyl	35
Chiwariidzo	Dimethoate	67
Aerodrome	Dimethoate	44
Shashi	Dimethoate	67
	Chlopyriphos- methyl	68
Magobo	Dimethoate	89

MATERIALS AND METHODS

Chemicals and materials

Acephate, dimethoate, monocrotophos, Malathion, Chlopyriphos- methyl standards (95 %) were provided by Kutsaga tobacco research Station, Harare, Reagents, ethyl acetate, acetone, sodium hydrogen carbonate, anhydrous sulphate were all analytical grades bought from SkyLabs, South Africa

Sample collection and storage

Cabbages were bought basing on place of origin. One cabbage was bought every day until day five. The five cabbages were then processed to make a composite sample. Samples were packed in separate polythene bags, sealed and labeled with a unique sample identity and placed in an ice chest box. All samples were brought to the lab and were stored at 5°C. These samples were then extracted and analyzed within 24hrs from the time of their collection.

Sample preparation

Fresh vegetable samples were thoroughly shredded and homogenized (for this only the edible portions were included, whereas bruised and/or rotten parts were removed). Twenty grams of the sample was macerated in 40ml of ethyl acetate. 5.0g of sodium hydrogen carbonate and 20.0g anhydrous sodium sulphate were added to remove moisture and further macerated for 3 minutes using the ultra-turax macerator (blender). The samples were then centrifuged for 5 minutes at 3,000 rpm to obtain the 2 phases. The supernatant was then transferred to a clean graduated cylinder (25ml) to measure its volume. This was then taken for clean-up and pre-concentration using solid phase extraction.

Solid phase extraction

A solid phase extraction was carried out using SPE columns. The florisil column (500mg/8ml) cartridge [13] was conditioned with 5 ml of a mixture solution of acetone: n-hexane (3:7, v/v) through the column. During the conditioning and sample loading steps, the sorbent was never allowed to dry. The extract column was fitted with 20-port vacuum manifold and a receiving flask was placed under the column to collect the eluate. The sample was loaded under vacuum at flow rates of 5ml min⁻¹. After extract had passed; the column was dried by vacuum aspiration. This was done under increased vacuum for a period of 30 minutes. The eluted mixture was then concentrated to 1 ml using a rotary evaporator (Buchi type) and then dried. This was then dissolved in 1 ml of ethyl acetate. Pesticides were then quantified using GC-NPD [14].

Gas Chromatography nitrogen phosphorus detection (GC-NPD) Analysis

GC (the Hewlett-Packard model 5890 II) equipped with an NPD detector with a split less injector at 250°C and the injection volume of 1 µL together with an auto sampler model 7673. A GLC capillary column-DB-

17 was used. The detector temperature was set at 280 °C with carrier gas and makeup gas as nitrogen with a 2.0 ml/min and 30 ml/min flow rate respectively, hydrogen at 5 ml/min and air at 80ml/min respectively employing the split less mode with temperature program, 100°C for 1 min, 100°C-150°C at 30°C/min, 150°C for 2 min, 150°C-205°C at 30°C/min, 205°C-260°C at 2°C/min, and 260°C for 1min [6]. Typical chromatogram for standard analytes obtained is shown in Fig 1 below. Analyte identification within the samples was based on chromatographic retention times i.e. if any of the targeted pesticides was found to match peak identity with standards e.g. Figs 1 and 2. Quantitation was based on peak area. Calibration curve method was used for quantitation. Regression equation for Acephate, dimethoate, monocrotophos, Malathion, Chlopyriphos- methyl were $y = 6.792x + 29376$, $R^2 = 0.994$, $y = 47.50x + 72257$, $R^2 = 0.998$, $y = 6.778x + 59736$, $R^2 = 0.994$, $y = 3.675x + 45912$, $R^2 = 0.996$, $y = 4.903x + 67754$, $R^2 = 0.994$ respectively. The limit of detection was 0.1 ng/ml while limit of quantification was 2.35 ng/ml. All tested percentage recoveries were above $76.20 \pm 4.53\%$.

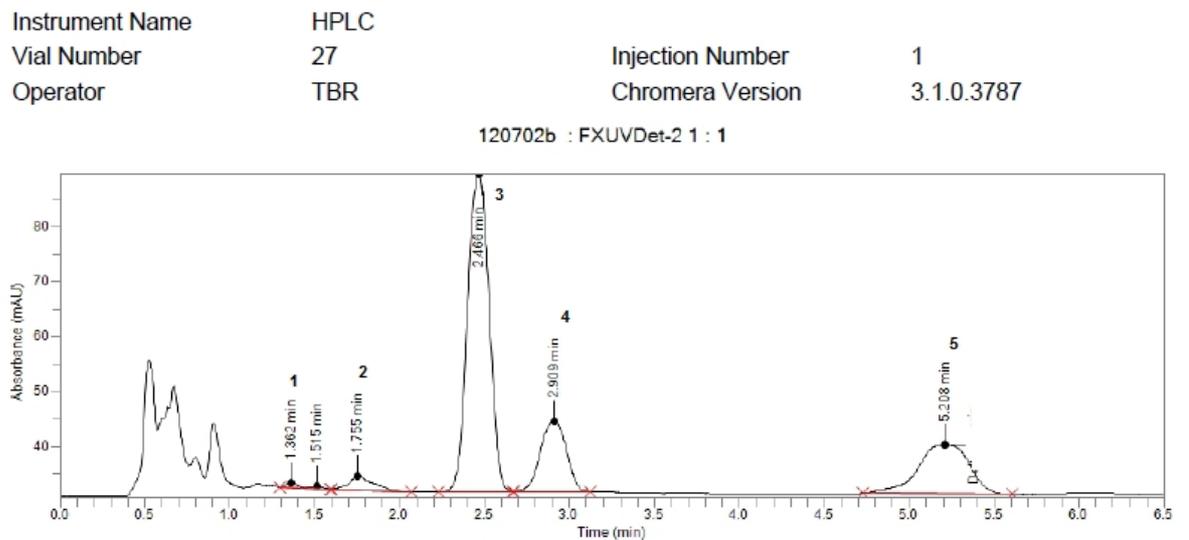


Fig 1. A typical GC chromatogram of standard pesticides, 1 = Acephate, 2 = dimethoate, 3 = Malathion, 4 = Chlopyriphos- methyl and 5 = monocrotophos

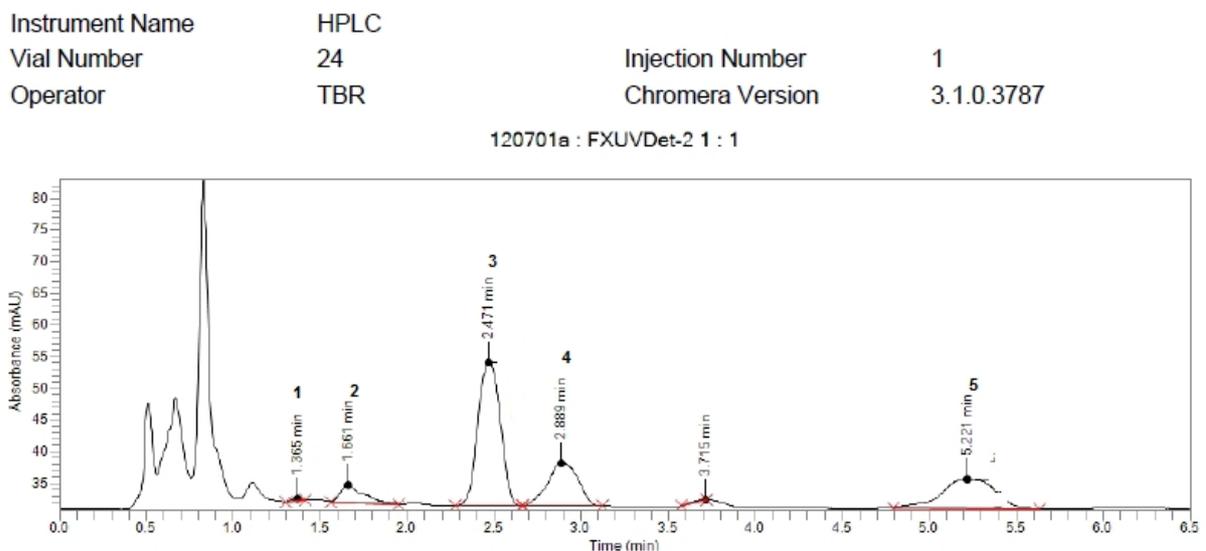


Fig 2. A GC chromatogram of a cabbage sample, 1 = Acephate, 2 = dimethoate, 3 = Malathion, 4 = Chlopyriphos- methyl and 5 = monocrotophos

RESULTS AND DISCUSSION

Figure 1 shows a typical chromatogram obtained for a cabbages sample. The concentration of OPPs in cabbage samples are shown in Table 2. Cabbages from Supa farming area consisted of all the OPPs while they were not detected in cabbages from Madziwa, Shamva and Shashi. Dimethoate was the mostly

detected pesticide. The highest level, 0.81 ± 0.02 was obtained from Aerodrome and the lowest level, 0.02 ± 0.00 was recorded for cabbages from Magobo. A total of 51 residues were detected in 180 samples corresponding to detection rate of 28%. The implicated organophosphate pesticides include; acephate, dimethoate; malathion, chlopyrifos- methyl and monocrotophos. Although organophosphate pesticides residues were detected in some vegetables they were far below the accepted maximum residual limits [16] Low levels obtained in this study imply that famers are sticking to stipulated times before they harvest their crops. Frequent detection of all pesticides in the super cabbage samples is worrisome as this may have health implications through synergistic tendencies [17]. Effects of long term exposure to these low levels of pesticides is usual is not known [17, 18].

Table 2. Concentration of Organophosphates in cabbage samples

Sample Origin	Concentration (mg/kg FW) of				
	Acephate	Dimethoate	Malathion	Chlopyrifos-methyl	monocrotophos
Musana	ND	0.33 ± 0.02	ND	ND	ND
Manhenga	ND	0.41 ± 0.01	0.06 ± 0.00	ND	ND
Supa	0.07 ± 0.00	0.21 ± 0.01	0.10 ± 0.02	0.19 ± 0.01	0.12 ± 0.03
Madziwa	ND	ND	ND	ND	ND
Glendale	ND	0.08 ± 0.00	ND	ND	ND
Matepatepa	ND	0.57 ± 0.02	ND	ND	ND
Shamva	ND	ND	ND	ND	ND
Chipadze	ND	0.07 ± 0.00	0.03 ± 0.00	ND	ND
Chiwaridzo	ND	0.19 ± 0.02	ND	ND	ND
Aerodrome	ND	0.81 ± 0.02	ND	ND	ND
Shashi	ND	ND	ND	ND	ND
Magobo	ND	0.33 ± 0.04	0.07 ± 0.00	0.02 ± 0.00	ND

FW = fresh weight, ND = not detected

CONCLUSION

All the analyzed cabbage samples consisted of levels that were below the recommended maximum residual limits implying that famers stuck to prescribed times before their cabbages could be harvested for human consumption however presence of low levels of all pesticide residues in some samples is worrisome since effects of chronic exposure to this low levels are not known especially on young children.

ACKNOWLEDGEMENT

Authors would like to thank Kutsaga Tobacco research station for their kind help and Bindura University of Science Education for providing block research funds.

REFERENCES

1. Codex Alimentarius Commission (2000). Food Standards Programme. Pesticide residues in food. Methods of analysis and sampling. Volume 2A Part 1. WHO
2. Zhou, L., Bai, Y., Wang, J. (2006). Organophosphorus pesticide residues in market foods in Shaanxi area, China. Food Chem. 98:240-242. FAO/WHO. Food and Agriculture
3. Waliszewski, SM., Carvajal, O., Infanzon, RM., Trujillo, P., Hart, MM (2004). Co partition ratios of persistent organochlorines pesticides between adipose tissue and blood serum lipids. Bull. Environ. Contam. Toxicol. 73:732-738
4. Wessels, D., Barr, DB., Mendola, P (2003). Use of Biomarkers to indicate exposure of children to organophosphate pesticides: Implications for a longitudinal study of children's environmental health, Environmental Health Perspectives, 111(16):1939-1946
5. Waliszewski, SM., Aguirre, AA., Infanzon RM., Carrillo LL., Sanchez, LT (2000). Comparison of Organochlorine pesticide levels in adipose tissue and blood serum from mothers living in Veracruz, Mexico. Bull. Environ. Contam. Toxicol. 64:8-15
6. Thrasher, JD., Heuser G., Broughton A (2002). Immunological abnormalities in humans chronically exposed to chlorpyrifos. Arch. Environ. Health. 57(3): 181-187
7. Pope, CN (1999). Organophosphorus pesticides: Do they all have the same mechanism of toxicity? J. Toxicol. Environ. Health. 2:161-181
8. Zahm, S (1992). Pesticides and non-hodgkins lymphoma. Cancer Res. 52:Suppl 19: 5485s-5488

9. Qio, D., Seidler, FJ., Tate, CA., Cousins, MM., Slotkin, TA (2003). Fetal chlorpyrifos exposure: adverse effects on brain cell development and cholinergic biomarkers emerge post nately and continue into adolescence and adulthood, *Environ. Health. Perspect.* 111(4): 536-544
10. Mukherjee, DP., Bhupander, K., Sanjay, K., Meenu, M., Gaur, R., Prakash, D., Singh, SK., Sharma, CS (2011). Occurrence and distribution of pesticide residues in selected seasona lvegetables from West Bengal. *Arch. Appl. Sci. Res.* 3(5):85-93
11. Park, El-Aty, Lee, Song, Shim, (2006). Residue Analysis of Organophosphorus and Organochlorine pesticides in Fruits and vegetables, *Journal of food health*, 1: 128-135
12. Wen Li, Ming Sun and Minzan Li(2013) A Survey of Determination for organophosphorus pesticide residue in Agricultural products, *Advanced Journal of Food Science and Technology* , China 5(4) pg. 381-386
13. Sajjad, AB., Niaz, AA., Muhammad, A., Muhammad, R A(2009). Determination of theorganophosphorus pesticide in vegetables by HPLC, *Journal of Agricultural and environmental science* 6(5) : 513-519.
14. Dasika, E., Tangirala, KT., Naishadam (2011). Survey on organophosphorus pesticide residues in human blood using HPLC. *J. Chromatogr. A* 743(1): 465-500
15. ASEAN (2007). Harmonization of Maximum Residue Limits (MRLs) of pesticides invegetables, *Crops publication* Serial no 1: 1-25
16. Terry, AV ., Stone, JD (2003). Repeated exposures to sub threshold doses of chlorpyrifos in rats: hippocampal damage, impaired axonal transport, and deficits in spatial learning. *J. Pharmacol. Exp. Theor.* 305(1): 375-84.
17. Farag, AT., El Okazy, AM., El Aswed AF (2003). Developmental toxicity study of chlorpyrifos in rats. *Reprod. Toxicol.* 17(2): 203-208

Citation of this article

Shasha D, Gwezere W, Dzomba P, Chayamiti T. Determination of Organophosphorus Pesticide Residues in Cabbages from Bindura Market Place by Solid Phase Extraction and Gas Liquid Chromatography. *Bull. Env. Pharmacol. Life Sci.*, Vol 3 (3) February 2014: 95-99