Bulletin of Environment, Pharmacology and Life Sciences Bull. Env. Pharmacol. Life Sci., Vol 12 [10] September 2023 :366-376 ©2023 Academy for Environment and Life Sciences, India Online ISSN 2277-1808 Journal's URL:http://www.bepls.com CODEN: BEPLAD REVIEW ARTICLE



Probiotic Proceedings of Lactobacillus and Bifidobacterium on Individual Vigor Remuneration: Contemporary Standing

Mithilesh Jaiswal¹, Kajol Batta², Juhi Sharma^{3*}, Shyamji Shukla⁴ and Rita Sharma⁵

¹Syncare Biotech Pvt. Ltd., Gwalior, (India)
 ^{2*}Department of Food Technology, ITM University, Gwalior (India)
 ³Jaipur National University, Jaipur (India).
 ⁴Department of Biotechnology, Mata Gujri Mahila Mahavidyalaya, Jabalpur (India).
 ⁵Department of Biotechnology, ITM University Gwalior (India)
 drmithileshjaiswal@gmail.com

ABSTRACT

During the phase of COVID 19, people of whole world were facing the biggest challenges of health problems. Everyone looking for the better options to enhance the immunity of the body to protect themselves from various viral and bacterial infections. Several nutritional supplements and functional foods are available which enhance the immunity of the body simultaneously fulfil the nutritional requirements. But these are insufficient to make body fit and ready to fight against outer enemies, microbial infections and several health problems. Probiotic is established to fill this gap as it has group of live beneficial microbes which provides health benefits to the hosts and protect them from microbial infections and various health problems. One of the most significant aspects of probiotics is that they have proven to be safe, cheap and capable of hindering with microbial infection. Probiotic also play a pivotal role in keeping up the resistance and the metabolic homeostasis effective to protect against pathogenic microbes. Use of probiotics is rapidly expanding with the globalization of functional food and health supplements. Various probiotic products are available containing different specific probiotic strains for specific health problems. Beneficial role of Probiotic microbes is strain specific and all the probiotic strains are lies under the genera of Lactobacillus, Bifidobacterium, Bacillus, Streptococcus, Enterococcus, Lactococcus, Leuconostoc, Saccharomyces. But most of the probiotic strains lies under the genera of Lactobacillus not fue probiotic strains of Lactobacillus not protect strains of Lactobacillus and Bifidobacterium in various health problems.

Key words: Probiotics, Synbiotic, Lactobacillus, Bifidobacterium, Lactic acid bacteria, Human health

Received 23.07.2023

Revised 21.08.2023

Accepted 21.09.2023

INTRODUCTION

In 1953, the German Scientist Werner Kollath was introduced the term probiotics by which to mean "active substances essential for a healthy life" [1]. FAO/WHO in 2002 given the world wide accepted definition of probiotic that live microorganism which when administered in adequate amount confer health benefits to the host. All probiotic strains come under the category of Lactic acid bacillus as all are have characteristic to produce lactic acid by the fermentation of carbohydrate sugar. Based on certain health promoting properties such as resistance to enteric pathogens, aid in lactose digestion, modulation of immune system and decreasing detoxification/excretion of toxic microbial metabolites [2], probiotics have emerged as the chief nutritional factor and laid down new opportunities for the food and nutrition field for scientists to improve food quality and develop new products with specific health benefits for different sub populations [3]. More specifically probiotics are active microorganism which benefit the host by improving the health of gastrointestinal tract through establishment of healthy microflora [4]

Probiotics have been investigated to determine potential beneficial effects in the prevention and treatment of a wide variety of systemic conditions. These conditions include rebalancing the population of bacteria in the gut, aiding digestion, inflammatory and autoimmune diseases such as rheumatoid arthritis, ulcerative colitis, multiple sclerosis, and hepatic encephalopathy [5]. Probiotics are considered as boon to improve the Gut health by colonizing healthy bacteria capable of producing various vitamins and essential metabolites for normal growth and development.

The most important characteristic of probiotic strains is that they generally considered to be safe for human consumption. Therefore, WHO considered maximum probiotic strains as GRAS (generally recognized as safe). However, various regulatory bodies from different countries specify some important parameters or characteristics for the potential strains which need to be tested before its commercialization or consumption. To be used as a probiotic, the selected strain must be able to tolerate gastrointestinal conditions, mucosal adhesions, and deprivation of competition [6]. Additionally, they should possess the abilities such as pathogen exclusion, anti-oxidant, antimicrobial, immuno-modulatory and food fermentation. Probiotics are also reported to be useful in modulating Human health [7, 8]. Most commonly micro-organisms used as probiotics belong to the heterogeneous group of lactic acid bacteria (genera Lactobacillus and Enterococcus) and the genus Bifidobacterium. Each group involves different species (*Lactobacillus acidophilus, Bifidobacterium bifidum*, etc.), including different strains [9].

Potential strains of Lactobacillus and Bifidobacterium, have been widely used as effective probiotics [10]. Lactobacillus and Bifidobacterium have been suggested to be associated with alleviation of lactose intolerance; prevention and cure of diarrhoea caused by viral, bacterial, and antibiotic or radiotherapy, immunomodulation; antimutagenic and anticarcinogenic effects and physiological and metabolic changes like reduce the body fat 14, blood cholesterol [11, 12].

There are many sources of exposure to Lactobacilli and Bifidobacteria. These sources include probiotics, fermented foodstuffs (e.g., yogurt, cheese, sauerkraut, Sourdough bread, Pickles, Beer, wine, cider and many other fermented foods), as well as the host's own microflora. In many traditional foods, such bacteria play an important role in preventing spoilage and the growth of pathogenic microorganisms. Some probiotic products that contain lactobacilli or bifidobacteria have long histories of safe use in some cases, for many decades [13]. They inhibit the growth of other harmful bacteria with lowering the pH by producing lactic acid and preserve the nutritive qualities of raw food material for an extended shelf life [14-16]. In healthy humans, lactobacilli are normally present in the oral cavity (103 –104cfu/g), the ileum (103 –107cfu/g), and the colon (104 -108cfu/g), and they are the dominant microorganism in the vagina. Increasing consumption of probiotic lactobacilli and *bifidobacteria* has not led to opportunistic infections in consumer [18]. However, infection due to lactobacilli and *bifidobacteria* are extremely rare and are estimated to represent 0.05%–0.4% of cases of infective endocarditis and bacteremia [17]. Lactobacilli constitute an integral part of the healthy gastrointestinal (GI) microecology and are involved in the host metabolism [19]. Due to fast growing awareness about the benefits of probiotic among the consumers, people looking for different probiotic products and health supplements to get health benefits and to boost their immunity to fight against different health problems. Therefore, various synbiotic (Probiotic and Prebiotic) products containing different strains of Bifidobacterium and Lactobacilli are introduced in the market by the probiotic manufactures for different health application. Furthermore, the growth and market size of probiotic products also increases day by day. Probiotics have widely been used in food, dairy, Pharmaceuticals, Nutraceuticals and fermentation as well as non-pharmacological sectors [20].

General Characteristic:

The most commonly used strains of Lactobacilli spp are Lactobacillus acidophilus, Lacticaseibacillus rhamnosus, Lacticaseibacillus casei, Lacticaseibacillus paracasei, Lactiplentibacillus plantarum, Limosilactobacillus fermentum, Limosilactobacillus reuteri, Levilactobacillus breve, Lactobacillus gasseri, Lactobacillus helveticus, Lactobacillus johnsonii and many more. Similarly, the most common strains of Bifidobacterium spp. are Bifidobacterium bifidum, Bifidobacterium longum, Bifidobacterium brevis, Bifidobacterium animalis, Bifidobacterium infantis etc.

Lactobacillus are rod shaped, gram positive, catalase negative, non- motile, oxidase negative, endospore forming, fermentative, facultative anaerobic or microaerophilic bacteria [21-23]. In most cases they form chains of varying length. Lactobacilli have a generation time from 25 to several hundred minutes. In humans and animals, they are found in the intestinal track and perform many beneficial functions including immunomodulation, suppression of enteric pathogens and maintenance of intestinal flora. Species of *Lactobacilli* are mostly homofermentative, but some are heterofermentative. The genus has been divided into three major subgroups and over 70 species are recognized [24].

Bifidobacterium are gram-positive, non-spore forming, anaerobic, pleomorphic bacteria [25-28]. *Bifidobacteria* have been shown to represent one of the most abundant genera present in a healthy gut early in life, being the most abundant genus present in the intestine of healthy breastfed infants, and to play an important role in gut homeostasis and immune system development [29-32].

Some Common health benefits of strains of Lactobacilli and Bifidobacterium spp. are:

- They can help to improve the health problems related to heart.
- They can also help to provide relieve in the problem of lactose intolerance.

- They are helpful in various food and skin allergies.
- They can reduce inflammation in the vagina due to an overgrowth of bacteria.
- They can improve unpleasant odor in exhaled breath.
- They help to reduce the frequency and duration of diarrhea.

Some best probiotic strains of Lactobacilli and Bifidobacterium for health benefits include:

- *Lactobacillus plantarum* (*Lactiplantibacillus plantarum*) inhibits the growth of harmful bacteria. They stimulate the digestive system, fights off disease-causing bacteria, and helps the body to produce vitamins.
- Lactobacillus acidophilus regulates stomach acidity levels.
- *Lactobacillus paracasei (Lacticaseibacillus paracasei)* strengthens the immune system.
- *Lactobacillus fermentum (Limosilactobacillus fermentum*) strengthens immune system and prevents gastrointestinal and upper respiratory infections
- *Lactobacillus reuteri* (*Limosilactobacillus reuteri*) supports heart health by balancing cholesterol levels. It also reduces ulcer-causing bacteria and supports female urinary tract and vaginal health.
- *Lactobacillus delbrueckii* subsp.*bulgaricus* is supports good digestion, prevents diarrhoea, and helps relieve symptoms of irritable bowel syndrome (IBS).
- *Lactobacillus rhamnosus (Lacticaseibacillus rhamnosus) is* naturally found in your gut, although you can eat foods or take supplements to increase its benefits. It's helpful in relieving IBS symptoms, treating diarrhoea, strengthening your gut health, and protecting against cavities.
- *Bifidobacterium longum ssp. Infantis* is commonly used to treat bowel problems, eczema, vaginal yeast infections, lactose intolerance, and urinary tract infections.
- *Bifidobacterium bifidum* can help manage your digestive system, improve IBS, and boost your immune system.
- *Bifidobacterium animalis ssp. Lactis* helps prevent infection. It helps to break down carbohydrates and synthesize vitamins.

Different beneficial roles of some important strains of Bifidobacterium and Lactobacilli are included in **Table 1**.

	Probiotic effect on	Dosage and	Function	Reference		
	Culture/ combination used	duration				
1	As neuroprotectants					
	L. acidophilus, L. casei, B.	2 X 109 CFU/g	Improved cognitive function and suppress	[31]		
	bifidumand L. fermentum	each for 12	inflammation			
		weeks.				
2	Combating stress and depression	on				
	L.acidophilus, L. rhamnosus, L.	50 billion for	Reduce detrimental effects of stress on	[32]		
	paracasi, L.plantarum, L.returi,	28 days	cognition			
	B. longum, B.lactis					
3	Skin problems					
	B.lactsB.longumand L.casei	1 X 10 ⁹ CFU/g	Effective in reducing SCORAD index and	[33]		
		each for 12	reducing the use of topical steroids in			
		weeks.	patients with moderate AD.			
4	Rheumatoid arthritis					
	L. acidophilus, L. casei,	2 × 109 CFU, for	Improved Disease Activity Score of 28	[34]		
	Bifidobacterium bifidum	8 weeks.	joints , insulin levels, and hs-CRP levels			
5	Anticancer agents					
	L.plantarum	1011 CFU, for 16	Probiotics decreased the serum zo nulin	[35]		
	L.acidophilus	days	concentration, duration of postoperative			
	and B.longum		pyrexia, inhibite the p38 MAP kinase			
	0		signaling pathway.			
6	Metabolic abnormality					
	Probiotic capsule:	10 ¹⁰ CFU, 12	Reduce the lipid levels and hs-CRP levels	[36]		
	L. acidophilus, B	week				
	bifidum, L. casei, L. fementum					

Table 1: Some best strains of Lactobacilli and Bifidobacterium which modulate the human health

	L. acidophilus, L. casei, L. rhamnosus, L. bulgaricus, Bifidobacterium breve, B. longum,	2X10 ¹¹ CFU, 6 week	Reduce the Blood glucose and HbA1c levels	[37]	
7	Allergies				
	L. rhamnosus	2×10 ¹⁰ CFU,for 6 months	Effective in inducing possible sustained unresponsiveness and modulation of the immune response.	[38]	
8	Respiratory tract infections				
	L. rhamnosus	2 X 10ºCFU, For 60 day	Reduce the No. of days in episodes with fever, diarrhea, and respiratory Illness	[39]	
	L.rhamnosus B.breve P.shermanii	2 X 10 ⁹ CFU for 6 month		[40]	

Health benefits and superiority of Lactobacilli and Bifidobacterium strains over other Strains of Lactic Acid Bacteria:

The digestive system as well as mucous membranes constitute natural sites of Bifidobacterium, whereas Lactobacillus bacteria inhibit the digestive and urogenital systems. *Bifidobacteria* are among the first colonisers of the sterile gastrointestinal tract of new born during breast feeding. Most strains of Lactobacilli can ferment amygdalin, cellobiose, fructose, galactose, glucose, lactose, maltose, mannose, salicin, sucrose and trehalose [41-45]. The genus of Bifidobacterium may, with the assistance of intracellular enzymes, break down polysaccharides which undergo conversion into glucose and fructose phosphates.

It was reported that Lactobacillus and Bifidobacillus containing probiotics were found to improve outcomes in acute infectious diseases outside of the gastrointestinal tract, such as upper and lower respiratory tract illnesses in infants and college students [46, 47]. It was also reported that the addition of fermenting *Lacticaseibacillus paracasei* to milk or rice milk resulted in reduced episodes of gastroenteritis, rhinitis, otitis, laryngitis, and tracheitis [48]. These finding suggested that the benefits to the host extend beyond local interactions in the intestinal tract between the gut organisms, enterocytes, and the immune system, perhaps involving microbial metabolites and/or migrating dendritic cells that reach distant locations such as the spleen and lymph nodes.

Lactobacilli and Bifidobacterium spp have been found to control intestinal disorders through a number of mechanisms to prevent destructive or pathogenic bacteria from mounting on and attaching to the intestinal epithelium: viz production and secretion of antimicrobial agents such as bacteriocins and organic acids [49, 50]. Bacteriocins are small proteins that have properties such as antitumour and anticholesterol activity which constitute a heterologous sub group of ribosomally synthesized antimicrobial peptides [51-53]. Bacteriocins have been reported to permeate the outer membrane and to induce the inactivation of Gramnegative bacteria in conjunction with other enhancing antimicrobial environmental factors, such as low temperature, organic acid and detergents [54, 55]. Some studies are also demonstrated that by products produced by *Lactobacillus spp* inhibited the growth of fungi such as *Aspergillus niger, Cladosporium herbarum and Penicillium verrucosum* in food materials [56, 57].

Bifidobacterium longum is the most frequently reported Bifidobacterium spp. having health benefits include competing with harmful bacteria for nutrients, colonization numbers and adherence to the intestinal epithelium. Control of epithelial cell proliferation and differentiation and homeostatic regulation of the immune system are tropic functions [58-60]. *Bifidobacteria* expand the number of mucosal lymphocytes, the size of germinal canters in lymphoid follicles and there is some research indicating a role in the induction of regulatory T cells in lymph follicles [61].

Mechanism of Action:

Mechanism of probiotic can be described as stimulation of phagocytic activity, complement-mediated bacterial killing, and immunoglobulin production [62, 63].

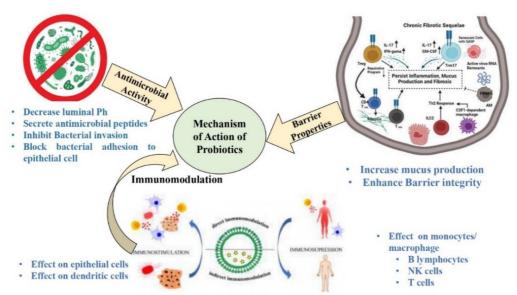


Figure 1: Mechanism of action of probiotics

Paraprobiotics and Postbiotics properties of Lactobacilli:

Parabiotics can be well described administration of small amount of inactivated microbial cells or cell fractions which can deliver health benefit to host [59]. On the other hand, the soluble products generated as a result of metabolism of probiotics are termed as postbiotics. These products have also been reported to confer health benefits to host [64].

Both paraprobiotics and postbiotics can efficiently be extracted from specific probiotic bacteria using techniques such as, thermal treatment, enzymatic treatments, solvent extraction, radiation (ionizing and UV rays) and many more [60, 61, 34] Postbiotics consist of secreted proteins and peptides, bacteriocins, organic acids [65]. Both endure health promoting properties such as immunomodulatory, anti-tumor, barrier-preservation, and antimicrobial properties [66]. Paraprobiotics comprises of peptidoglycans, surface proteins, cell wall polysaccharides. These properties of Lactobacilli are included in **Table No.2**.

Table 2: Paraprobiotic of some Lactobacilli strains and health benefits:						
Probiotic	Active component	Specificity	Beneficial effect		References	
			Strain	Effect		
Lactobacilli	ctobacilli Peptidoglycan Polymer of repeating units of N- acetylglucosamine and N- acetylmuramic disaccharide	repeating units of N- acetylglucosamine and N-	Lactobacillus casei, Lactobacillus johnsonii, Lactobacillus plantarum	Suppress interleukin-12 (IL-12) production, preventing inflammatory bowel diseases	[66-69]	
		Lactobacillus salivarius	Anti-inflammatory properties	-		
		Lactobacillus rhamnosus	Boast Innate and systemic adaptive immune responses in mice			
	Teichoic Acid	Covalently linked to peptidoglycan	<i>L. plantarum</i> LTA	Anti-inflammatory effects and immunomodulatory properties	[70]	
	Cell-Wall Polysaccharides	Exopolysaccharides providing the interaction of the	L. rhamnosus	Immuno-suppressive effect	[71, 72]	
	bacteria with the environment	L. plantarum	anti-inflammatory effect, response against enterotoxigenic,			
		Lactobacillus delbrueckii	Anti-viral property			

		Lactobacillus acidophilus	Effective against colon cancer	
Cell Surface Proteins	e Attached to cell surfa	Attached to cell surface via covalent or non-covalent linkage		
LPXTG Proteins	Comprising of LPXTG signal and forms linkage with cell wall via sortase A (SrtA)	<i>L. plantarum</i> and related species	Initiate bacteria-host interaction	[73-75]
S-Layer Proteins	Linked to peptidoglycan via non-covalent linkage	Lactobacillus paracasei subp. paracasei, L. rhamnosus, L. casei	Prevent the infection of pathogenic bacteria Prevent the adherence of <i>Shigella sonnei</i>	[76]
		L. helveticus fb213, L. acidophilus fb116 and L.		
Moonlighting Proteins	assist the colonization of probiotic strains in intestinal tract	L. plantarum, Lactobacillus fermentum (L. fermentum), and L.	Prevention of pathogens in intestinal tract	[77]

Probiotic products containing potential strains of Lactobacilli and Bifidobacterium spp.:

Lactobacilli are often found in dairy products and other probiotic functional food. Lactobacillus delbrueckii subsp bulgaricus are used in the preparation of yogurt; Lactobacillus acidophilus is used in the preparation of acidophilus milk; Lactobacillus helveticus, as well as L. delbrueckii subsp bulgaricus, are used to make Swiss, Mozzarella, provolone, Romano, and parmesan cheeses etc. The lactobacilli are usually more resistant to acidic conditions than are other LAB, being able to grow at pH values as low as 4. This enables them to continue to grow during natural lactic fermentations when the pH has dropped too low for other LAB to grow, so they are often responsible for the final stages of many lactic acid fermentations. Bifidobacterium spp is an extremely versatile strain found in foods such as yogurt and fermented milk [78]. Apart from above, several species of probiotic are also used in the preparation of various Pharmaceuticals and Nutraceuticals probiotic and Synbiotic products which are available in the markets as a single culture or combination of multi strains in different forms like capsules. Sachets, dry syrup, tablets, gummies etc. [79, 80]. Synbiotic contains the benefits of both probiotics and prebiotics. Prebiotics are generally inactive ingredients and dietary fibres which are not easily digest by the body and it act as food for probiotic strains and helps them to grow. There are various prebiotics ingredients like Fructooligosaccharide, inulin etc. which are commonly used by the manufacture in the preparation of synbiotic products. Some common probiotic functional foods and probiotic strains are included in Table no. 3.

Table 3: Some Probiotic functional foods containing potential strains of Lactobacilli and Bifidobacterium:

Product	Probiotic Strain	Health benefits	References	
Kefir	Lactobacillus spp.	Anti-Hypertensive, Anti-Cancer, Anti-inflammatory	[81-83, 89]	
Sauerkraut	Lactobacillus plantarum, Lactobacillus brevis,	Anti-inflammatory, Anti-Cancer, Treats skin infections, Reduces oxidative stress	[84]	
Kimchi	Lactobacillus plantarum, and Lactobacillus brevis	anticancer, antiobesity, andanti-atherosclerotic	[85, 86]	
Pickles	Lactobacillus plantarum, and Lactobacillus pentosus	Antimutagenic, cholesterol-lowering effects	[85]	
Yoghurt	Lactobacillus plantarum 201 LRR	Improves digestion, Anticarcinogenic, anti-diabetic, Hypocholesterolemic effect, anti- diarrhoeal	[86, 87]	
Yakult	Lactobacillus spp. and Bifidobacterium spp		[88]	

Major players in the field of probiotics Market:

The probiotic market of is assigned into three categories based on product viz. probiotic food & beverages, probiotic dietary supplements, and animal feed probiotics. The major key players operating in this market comprises of BioGaia A.B, Danone, Chr. Hansen Holding A/S, Yakult Honsha Co. Ltd., Probi A B, Lifeway Foods, Inc., Nestle S.A., Ganeden, Inc., E. I. du Pont de Nemours and Company, and Protexin. Brands that exist in Indian probiotic industry are Nestle, Amul, Yakult Danone and Mother Dairy along with other minor players operating in different regions in their own capacities.

CONCLUSION

Most of the potential strains of probiotic lies under the genera of Lactobacilli and Bifidobacterium. It is well established that probiotics have impact on the immune system and this interaction is directly linked to gut microbes, their polysaccharide antigens, and key metabolites produced by probiotic strains. Gut microbiota modification with specific probiotic and Synbiotic products containing potential strains of Lactobacilli and Bifidobacterium spp. might offer a novel and cost-effective therapy to reduce the risk of number of diseases. Probiotic functional foods as well us dietary supplements containing potential strains of Bifidobacterium and Lactobacilli are capable to boost immunity as well as provides health benefits in several health problems. However, awareness needs to be created in general public for the selection and consumption of correct probiotic products to get health benefits. Also, future research needs in terms of the underlying mechanism of action involved in each of the observed effects. Advanced techniques such as encapsulation of probiotic bacteria can be undertaken to produce food products with beneficial effect.

ACKNOWLEDGMENT

The author received no financial support for the research.

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

Mithilesh Jaiswal: Writing – original draft, Visualization, Editing Kajol Batta- Writing – original draft, Editing and Revising Juhi Sharma: Visualization, Editing Shyamji Shukla: Supervision Rita Sharma: Editing and Revising

FUNDING INFORMATION

The authors received no financial support

REFERENCES

- 1. Gasbarrini, G., Bonvicini, F., Gramenzi, A. Probiotics History. J Clin Gastroenterol (2016). Nov/Dec;50 Suppl 2, Proceedings from the 8th Probiotics, Prebiotics & New Foods for Microbiota and Human Health meeting held in Rome, Italy on September 13-15, 2015:116-S119.
- 2. Nagpal, R., Kumar, A., Kumar, M., Behare, P. V., Jain, S., & Yadav, H. (2012). Probiotics, their health benefits and applications for developing healthier foods: a review. FEMS microbiology letters, 334(1), 1-15.
- 3. Markowiak, P., & Śliżewska, K. (2017). Effects of probiotics, prebiotics, and synbiotics on human health. Nutrients, 9(9), 1021.
- 4. Fuller, R. (1989). Probiotic in man and animals. J. Appl. Bacteriol., 66, 65-378.
- 5. Liu, Y., Alookaran, J. J., & Rhoads, J. M. (2018). Probiotics in autoimmune and inflammatory disorders. Nutrients, 10(10), 1537.
- 6. Marco, M. L., Heeney, D., Binda, S., Cifelli, C. J., Cotter, P. D., Foligné, B., ... & Hutkins, R. (2017). Health benefits of fermented foods: microbiota and beyond. Current opinion in biotechnology, 44, 94-102.
- 7. Holzapfel, W. H., & Schillinger, U. (2002). Introduction to pre-and probiotics. Food research international, 35(2-3), 109-116.
- 8. Bermudez-Brito, M., Plaza-Díaz, J., Muñoz-Quezada, S., Gómez-Llorente, C., & Gil, A. (2012). Probiotic mechanisms of action. Annals of Nutrition and Metabolism, 61(2), 160-174.
- 9. Fijan, S. (2014). Microorganisms with claimed probiotic properties: an overview of recent literature. International journal of environmental research and public health, 11(5), 4745-4767.
- 10. Gismondo, M. R., Drago, L., & Lombardi, A. (1999). Review of probiotics available to modify gastrointestinal flora. International journal of antimicrobial agents, 12(4), 287-292.

- 11. Raygan, F., Rezavandi, Z., Bahmani, F., Ostadmohammadi, V., Mansournia, M. A., Tajabadi-Ebrahimi, M., ... & Asemi, Z. (2018). The effects of probiotic supplementation on metabolic status in type 2 diabetic patients with coronary heart disease. Diabetology & metabolic syndrome, 10, 1-7.
- 12. Razmpoosh, E., Javadi, A., Ejtahed, H. S., Mirmiran, P., Javadi, M., & Yousefinejad, A. (2019). The effect of probiotic supplementation on glycemic control and lipid profile in patients with type 2 diabetes: A randomized placebo controlled trial. Diabetes & Metabolic Syndrome: Clinical Research & Reviews, 13(1), 175-182.
- 13. Rowland, I., Gibson, G., Heinken, A., Scott, K., Swann, J., Thiele, I., & Tuohy, K. (2018). Gut microbiota functions: metabolism of nutrients and other food components. European journal of nutrition, 57, 1-24.
- 14. Kohlmeier, M. (2015). Absorption, Transport, and Retention in Nutrient Metabolism. Elsevier. 37–93.
- 15. Gopal, P. K. (2011). Lactic Acid Bacteria: Lactobacillus spp.: Lactobacillus acidophilus. in Encyclopedia of Dairy Sciences: Second Edition Elsevier Inc. 91–95.
- 16. Lazar, V., Ditu, L. M., Pircalabioru, G. G., Gheorghe, I., Curutiu, C., Holban, A. M., ... & Chifiriuc, M. C. (2018). Aspects of gut microbiota and immune system interactions in infectious diseases, immunopathology, and cancer. Frontiers in immunology, 9, 18-30.
- 17. Borriello, S. P., Hammes, W. P., Holzapfel, W., Marteau, P., Schrezenmeir, J., Vaara, M., & Valtonen, V. (2003). Safety of probiotics that contain lactobacilli or bifidobacteria. *Clinical infectious diseases*, *36*(6), 775-780.
- 18. Zawistowska-Rojek, A., & Tyski, S. (2018). Are probiotic really safe for humans?. Polish journal of microbiology, 67(3), 251-258.
- 19. Walter, J. (2008). Ecological role of lactobacilli in the gastrointestinal tract: implications for fundamental and biomedical research. Applied and environmental microbiology, 74(16), 4985-4996.
- 20. Anandharaj M and Sivasankari B (2014): Isolation of potentialprobiotic lactobacillus oris HMI68 from mother's milkwith cholesterol-reducing property. *J Biosci Bioeng.* Vol. 118.153-159.
- 21. Corsetti, A., De Angelis, M., Dellaglio, F., Paparella, A., Fox, P. F., Settanni, L., & Gobbetti, M. (2003). Characterization of sourdough lactic acid bacteria based on genotypic and cell-wall protein analyses. Journal of Applied Microbiology, 94(4), 641-654.
- 22. Hamad, S. H., Dieng, M. C., Ehrmann, M. A., & Vogel, R. F. (1997). Characterization of the bacterial flora of Sudanese sorghum flour and sorghum sourdough. Journal of Applied Microbiology, 83(6), 764-770.
- 23. Gaglio, R., Alfonzo, A., Polizzotto, N., Corona, O., Francesca, N., Russo, G., ... & Settanni, L. (2018). Performances of different metabolic lactobacillus groups during the fermentation of pizza doughs processed from semolina. Fermentation, 4(3), 61.
- 24. Schleifer, K. H., & Ludwig, W. (1995). Phylogeny of the genus Lactobacillus and related genera. Systematic and Applied Microbiology, 18(4), 461-467.
- 25. Ventura, M., Canchaya, C., Tauch, A., Chandra, G., Fitzgerald, G. F., Chater, K. F., & Van Sinderen, D. (2007). Genomics of Actinobacteria: tracing the evolutionary history of an ancient phylum. Microbiology and molecular biology reviews, 71(3), 495-548.
- 26. Turroni, F., Foroni, E., Pizzetti, P., Giubellini, V., Ribbera, A., Merusi, P., ... & Ventura, M. (2009). Exploring the diversity of the bifidobacterial population in the human intestinal tract. Applied and environmental microbiology, 75(6), 1534-1545.
- 27. Arumugam, M., Raes, J., Pelletier, E., Le Paslier, D., Yamada, T., Mende, D. R., ... & Bork, P. (2011). Enterotypes of the human gut microbiome. nature, 473(7346), 174-180.
- 28. Stark, P. L., & Lee, A. (1982). The microbial ecology of the large bowel of breastfed and formula-fed infants during the first year of life. Journal of medical microbiology, 15(2), 189-203.
- 29. Penders, J., Thijs, C., Vink, C., Stelma, F. F., Snijders, B., Kummeling, I., ... & Stobberingh, E. E. (2006). Factors influencing the composition of the intestinal microbiota in early infancy. Pediatrics, 118(2), 511-521.
- 30. Benno, Y., Sawada, K., & Mitsuoka, T. (1984). The intestinal microflora of infants: composition of fecal flora in breast-fed and bottle-fed infants. Microbiology and immunology, 28(9), 975-986.
- 31. Akbari, E., Asemi, Z., Daneshvar Kakhaki, R., Bahmani, F., Kouchaki, E., Tamtaji, O. R., ... & Salami, M. (2016). Effect of probiotic supplementation on cognitive function and metabolic status in Alzheimer's disease: a randomized, double-blind and controlled trial. Frontiers in aging neuroscience, 8, 256.
- 32. Papalini, S., Michels, F., Kohn, N., Wegman, J., van Hemert, S., Roelofs, K., ... & Aarts, E. (2019). Stress matters: randomized controlled trial on the effect of probiotics on neurocognition. Neurobiology of stress, 10, 100141.
- Navarro-López, V., Ramírez-Boscá, A., Ramón-Vidal, D., Ruzafa-Costas, B., Genovés-Martínez, S., Chenoll-Cuadros, E., ... & Codoñer-Cortés, F. M. (2018). Effect of oral administration of a mixture of probiotic strains on SCORAD index and use of topical steroids in young patients with moderate atopic dermatitis: a randomized clinical trial. JAMA dermatology, 154(1), 37-43.
- 34. Zamani, B., Golkar, H. R., Farshbaf, S., Emadi-Baygi, M., Tajabadi-Ebrahimi, M., Jafari, P., ... & Asemi, Z. (2016). Clinical and metabolic response to probiotic supplementation in patients with rheumatoid arthritis: a randomized, double-blind, placebo-controlled trial. International journal of rheumatic diseases, 19(9), 869-879.
- 35. Liu, Z. H., Huang, M. J., Zhang, X. W., Wang, L., Huang, N. Q., Peng, H., ... & Wang, J. P. (2013). The effects of perioperative probiotic treatment on serum zonulin concentration and subsequent postoperative infectious complications after colorectal cancer surgery: a double-center and double-blind randomized clinical trial. The American journal of clinical nutrition, 97(1), 117-126.
- 36. Mohseni, S., Bayani, M., Bahmani, F., Tajabadi-Ebrahimi, M., Bayani, M. A., Jafari, P., & Asemi, Z. (2018). The beneficial effects of probiotic administration on wound healing and metabolic status in patients with diabetic foot

ulcer: a randomized, double-blind, placebo-controlled trial. Diabetes/Metabolism Research and Reviews, 34(3), e2970.

- 37. Tang, M. L., Ponsonby, A. L., Orsini, F., Tey, D., Robinson, M., Su, E. L., ... & Donath, S. (2015). Administration of a probiotic with peanut oral immunotherapy: a randomized trial. Journal of Allergy and Clinical Immunology, 135(3), 737-744.
- 38. Luoto, R., Ruuskanen, O., Waris, M., Kalliomäki, M., Salminen, S., & Isolauri, E. (2014). Prebiotic and probiotic supplementation prevents rhinovirus infections in preterm infants: a randomized, placebo-controlled trial. Journal of Allergy and Clinical Immunology, 133(2), 405-413.
- 39. Kukkonen, K., Savilahti, E., Haahtela, T., Juntunen-Backman, K., Korpela, R., Poussa, T., ... & Kuitunen, M. (2008). Long-term safety and impact on infection rates of postnatal probiotic and prebiotic (synbiotic) treatment: randomized, double-blind, placebo-controlled trial. Pediatrics, 122(1), 8-12.
- 40. Hoyles, L., Inganäs, E., Falsen, E., Drancourt, M., Weiss, N., McCartney, A. L., & Collins, M. D. (2002). Bifidobacterium scardovii sp. nov., from human sources. International Journal of Systematic and Evolutionary Microbiology, 52(3), 995-999.
- 41. Aron, N. M., Boev, M., & Bahrim, G. (2015). Probiotics and therapeutic effect in clinical practice–Review. Romanian Biotechnological Letters, 20(1), 10162-10175.
- 42. Okamoto, M., Benno, Y., Leung, K. P., & Maeda, N. (2008). Bifidobacterium tsurumiense sp. nov., from hamster dental plaque. International journal of systematic and evolutionary microbiology, 58(1), 144-148.
- 43. Ruiz, L., Delgado, S., Ruas-Madiedo, P., Sánchez, B., & Margolles, A. (2017). Bifidobacteria and their molecular communication with the immune system. Frontiers in microbiology, 8, 2345.
- 44. Modesto, M., Watanabe, K., Arita, M., Satti, M., Oki, K., Sciavilla, P., ... & Mattarelli, P. (2019). Bifidobacterium jacchi sp. nov., isolated from the faeces of a baby common marmoset (Callithrix jacchus). International journal of systematic and evolutionary microbiology, 69(8), 2477-2485.
- 45. Maldonado, J., Cañabate, F., Sempere, L., Vela, F., Sánchez, A. R., Narbona, E., ... & Lara-Villoslada, F. (2012). Human milk probiotic Lactobacillus fermentum CECT5716 reduces the incidence of gastrointestinal and upper respiratory tract infections in infants. Journal of pediatric gastroenterology and nutrition, 54(1), 55-61.
- 46. Rautava, S.; Salminen, S.; Isolauri, E. (2009): Specific probiotics in reducing the risk of acute infections in infancy— A randomised, double-blind, placebo-controlled study. *Br. J. Nutr.* Vol. 101.1722–1726.
- 47. Nocerino, R., Paparo, L., Terrin, G., Pezzella, V., Amoroso, A., Cosenza, L., ... & Canani, R. B. (2017). Cow's milk and rice fermented with Lactobacillus paracasei CBA L74 prevent infectious diseases in children: a randomized controlled trial. Clinical Nutrition, 36(1), 118-125.
- 48. Reid, G. (2001). Probiotic agents to protect the urogenital tract against infection. The American journal of clinical nutrition, 73(2), 437s-443s.
- 49. Rycroft, C. E., Fooks, L. J., & Gibson, G. R. (1999). Methods for assessing the potential of prebiotics and probiotics. Current Opinion in Clinical Nutrition & Metabolic Care, 2(6), 481-484.
- 50. Cotter, P. D., Ross, R. P., & Hill, C. (2013). Bacteriocins—a viable alternative to antibiotics?. Nature Reviews Microbiology, 11(2), 95-105.
- 51. Dobson, A., Cotter, P. D., Ross, R. P., & Hill, C. (2012). Bacteriocin production: a probiotic trait?. Applied and environmental microbiology, 78(1), 1-6.
- 52. Cleveland, J., Montville, T. J., Nes, I. F., & Chikindas, M. L. (2001). Bacteriocins: safe, natural antimicrobials for food preservation. International journal of food microbiology, 71(1), 1-20.
- 53. Chatterjee, S., Chatterjee, S., Lad, S. J., Phansalkar, M. S., Rupp, R. H., Ganguli, B. N., ... & KOGLER, H. (1992). Mersacidin, a new antibiotic from bacillus fermentation, isolation, purification and chemical characterization. The Journal of antibiotics, 45(6), 832-838.
- 54. Laitila, A., Alakomi, H. L., Raaska, L., Mattila-Sandholm, T., & Haikara, A. (2002). Antifungal activities of two Lactobacillus plantarum strains against Fusarium moulds in vitro and in malting of barley. Journal of applied microbiology, 93(4), 566-576.
- 55. Gerez, C. L., Torres, M. J., De Valdez, G. F., & Rollán, G. (2013). Control of spoilage fungi by lactic acid bacteria. Biological Control, 64(3), 231-237.
- 56. Alberoni, D., Gaggìa, F., Baffoni, L., Modesto, M. M., Biavati, B., & Di Gioia, D. (2019). Bifidobacterium xylocopae sp. nov. and Bifidobacterium aemilianum sp. nov., from the carpenter bee (Xylocopa violacea) digestive tract. Systematic and applied microbiology, 42(2), 205-216.
- 57. Duranti, S., Lugli, G. A., Milani, C., James, K., Mancabelli, L., Turroni, F., ... & Ventura, M. (2019). Bifidobacterium bifidum and the infant gut microbiota: an intriguing case of microbe-host co-evolution. Environmental microbiology, 21(10), 3683-3695.
- 58. Ringø, E., & Gatesoupe, F. J. (1998). Lactic acid bacteria in fish: a review. Aquaculture, 160(3-4), 177-203.
- 59. Vijayaram, S., & Kannan, S. (2014). Studies on probiotic analysis in the gut region of fresh water fishes in periyar lake, Kerala. Int J Pharm Sci Res Rev, 3, 17-20.
- 60. Taverniti, V., & Guglielmetti, S. (2011). The immunomodulatory properties of probiotic microorganisms beyond their viability (ghost probiotics: proposal of paraprobiotic concept). Genes & nutrition, 6(3), 261-274.
- 61. Aguilar-Toalá, J. E., Hall, F. G., Urbizo-Reyes, U. C., Garcia, H. S., Vallejo-Cordoba, B., González-Córdova, A. F., ... & Liceaga, A. M. (2020). In silico prediction and in vitro assessment of multifunctional properties of postbiotics obtained from two probiotic bacteria. Probiotics and antimicrobial proteins, 12, 608-622.

- Kamiya, T., Wang, L., Forsythe, P., Goettsche, G., Mao, Y., Wang, Y., ... & Bienenstock, J. (2006). Inhibitory effects of Lactobacillus reuteri on visceral pain induced by colorectal distension in Sprague-Dawley rats. Gut, 55(2), 191-196.
- 63. Dinić, M., Lukić, J., Djokić, J., Milenković, M., Strahinić, I., Golić, N., & Begović, J. (2017). Lactobacillus fermentum postbiotic-induced autophagy as potential approach for treatment of acetaminophen hepatotoxicity. Frontiers in microbiology, 8, 594.
- 64. Nakamura, F., Ishida, Y., Sawada, D., Ashida, N., Sugawara, T., Sakai, M., ... & Fujiwara, S. (2016). Fragmented lactic acid bacterial cells activate peroxisome proliferator-activated receptors and ameliorate dyslipidemia in obese mice. Journal of agricultural and food chemistry, 64(12), 2549-2559.
- 65. Shin, H. S., Park, S. Y., Lee, D. K., Kim, S. A., An, H. M., Kim, J. R., ... & Ha, N. J. (2010). Hypocholesterolemic effect of sonication-killed Bifidobacterium longum isolated from healthy adult Koreans in high cholesterol fed rats. Archives of pharmacal research, 33, 1425-1431.
- 66. Lebeer, S., Vanderleyden, J., & De Keersmaecker, S. C. (2010). Host interactions of probiotic bacterial surface molecules: comparison with commensals and pathogens. Nature Reviews Microbiology, 8(3), 171-184.
- 67. Shida, K., Kiyoshima-Shibata, J., Kaji, R., Nagaoka, M., & Nanno, M. (2009). Peptidoglycan from lactobacilli inhibits interleukin-12 production by macrophages induced by Lactobacillus casei through Toll-like receptor 2-dependent and independent mechanisms. Immunology, 128(1pt2), e858-e869.
- 68. Fernandez, E. M., Valenti, V., Rockel, C., Hermann, C., Pot, B., Boneca, I. G., & Grangette, C. (2011). Anti-inflammatory capacity of selected lactobacilli in experimental colitis is driven by NOD2-mediated recognition of a specific peptidoglycan-derived muropeptide. Gut, 60(8), 1050-1059.
- 69. Kolling, Y., Salva, S., Villena, J., & Alvarez, S. (2018). Are the immunomodulatory properties of Lactobacillus rhamnosus CRL1505 peptidoglycan common for all Lactobacilli during respiratory infection in malnourished mice?. PLoS One, 13(3), e0194034.
- Mohamadzadeh, M., Pfeiler, E. A., Brown, J. B., Zadeh, M., Gramarossa, M., Managlia, E., ... & Klaenhammer, T. R. (2011). Regulation of induced colonic inflammation by Lactobacillus acidophilus deficient in lipoteichoic acid. Proceedings of the National Academy of Sciences, 108(supplement_1), 4623-4630.
- 71. Bleau, C. 1., Monges, A., Rashidan, K., Laverdure, J. P., Lacroix, M., Van Calsteren, M. R., ... & Lamontagne, L. (2010). Intermediate chains of exopolysaccharides from Lactobacillus rhamnosus RW-9595M increase IL-10 production by macrophages. Journal of applied microbiology, 108(2), 666-675.
- 72. Nikolic, M., López, P., Strahinic, I., Suárez, A., Kojic, M., Fernández-García, M., ... & Ruas-Madiedo, P. (2012). Characterisation of the exopolysaccharide (EPS)-producing Lactobacillus paraplantarum BGCG11 and its non-EPS producing derivative strains as potential probiotics. International Journal of Food Microbiology, 158(2), 155-162.
- 73. Jensen, H., Roos, S., Jonsson, H., Rud, I., Grimmer, S., van Pijkeren, J. P., ... & Axelsson, L. (2014). Role of Lactobacillus reuteri cell and mucus-binding protein A (CmbA) in adhesion to intestinal epithelial cells and mucus in vitro. Microbiology, 160(Pt 4), 671.
- 74. Sengupta, R., Altermann, E., Anderson, R. C., McNabb, W. C., Moughan, P. J., & Roy, N. C. (2013). The role of cell surface architecture of lactobacilli in host-microbe interactions in the gastrointestinal tract. Mediators of inflammation, 2013.
- 75. Zhang, B., Zuo, F., Yu, R., Zeng, Z., Ma, H., & Chen, S. (2015). Comparative genome-based identification of a cell wallanchored protein from Lactobacillus plantarum increases adhesion of Lactococcus lactis to human epithelial cells. Scientific reports, 5(1), 14109.
- 76. Zhang, Y. C., Zhang, L. W., Tuo, Y. F., Guo, C. F., Yi, H. X., Li, J. Y., ... & Du, M. (2010). Inhibition of Shigella sonnei adherence to HT-29 cells by lactobacilli from Chinese fermented food and preliminary characterization of S-layer protein involvement. Research in microbiology, 161(8), 667-672.
- 77. Spurbeck, R. R., & Arvidson, C. G. (2010). Lactobacillus jensenii surface-associated proteins inhibit Neisseria gonorrhoeae adherence to epithelial cells. Infection and immunity, 78(7), 3103-3111.
- Granato, D., Branco, G. F., Nazzaro, F., Cruz, A. G., & Faria, J. A. (2010). Functional foods and nondairy probiotic food development: trends, concepts, and products. Comprehensive reviews in food science and food safety, 9(3), 292-302.
- 79. Min, M., Bunt, C. R., Mason, S. L., & Hussain, M. A. (2019). Non-dairy probiotic food products: An emerging group of functional foods. Critical reviews in food science and nutrition, 59(16), 2626-2641.
- 80. Simova, E., Beshkova, D., Angelov, A., Hristozova, T. S., Frengova, G., & Spasov, Z. (2002). Lactic acid bacteria and yeasts in kefir grains and kefir made from them. Journal of Industrial Microbiology and Biotechnology, 28, 1-6.
- 81. Witthuhn, R. C., Schoeman, T., & Britz, T. J. (2004). Isolation and characterization of the microbial population of different South African kefir grains. International Journal of Dairy Technology, 57(1), 33-37.
- 82. Chen, H. C., Wang, S. Y., & Chen, M. J. (2008). Microbiological study of lactic acid bacteria in kefir grains by culturedependent and culture-independent methods. Food microbiology, 25(3), 492-501.
- 83. Jayakumar, B.D., Kontham, K.V., Kesavan, M.N., Bindhumol, I., Pandey, A. (2012): Probiotic fermented food for human benefits. *Eng. Life Sci.* Vol.4. 377-390.
- 84. Raak, C., Ostermann, T., Boehm, K., & Molsberger, F. (2014). Regular consumption of sauerkraut and its effect on human health: a bibliometric analysis. Global advances in health and medicine, 3(6), 12-18.
- 85. Park, K. Y., & Kim, B. K. (2011). Lactic acid bacteria in vegetable fermentations. Lactic Acid Bacteria, Microbial and Functional Aspects, 4th ed.; Lahtinen, S., Ouwehand, AC, Salminen, S., Von Wright, A., Eds, 187-211.

- 86. Capela, P., Hay, T. K. C., & Shah, N. P. (2006). Effect of cryoprotectants, prebiotics and microencapsulation on survival of probiotic organisms in yoghurt and freeze-dried yoghurt. Food Research International, 39(2), 203-211.
- 87. Banerjee, U., Malida, R., Panda, R., Halder, T., & Roymahapatra, G. (2017). Variety of yogurt and its health aspectsa brief review. International Journal of Innovative Practice and Applied Research (IJIPAR).
- 88. Chuayana, J. E., Ponce, C. V., Rivera, M. R. B., & Cabrera, E. C. (2003). Antimicrobial activity of probiotics from milk products. Phil J. Microbiol. Infect. Dis, 32(2), 71-74.
 89. Shenderov, B. A. (2013). Metabiotics: novel idea or natural development of probiotic conception. Microbial ecology
- in Health and Disease, 24(1), 20399.

CITATION OF THIS ARTICLE

Mithilesh J, Kajol B, Juhi S, Shyamji S and Rita S Probiotic Proceedings of Lactobacillus and Bifidobacterium on Individual Vigor Remuneration: Contemporary Standing. Bull. Env.Pharmacol. Life Sci., Vol 12 [10] September 2023: 366-376